

## ORIGINAL RESEARCH

# Prognostic Impact of P16 and EGFR Overexpression in Patients with Cervical Cancer

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### ABSTRACT

**Background:** Cervical cancer is the second most prevalent cancer, both in terms of incidence and mortality, after breast cancer in regions with lower Human Development Index (HDI). However, it holds the distinction of being the most frequently diagnosed cancer in 28 countries and the primary cause of cancer-related deaths in 42 countries. The majority of these countries are located in Sub-Saharan Africa and South Eastern Asia. Cervical cancer stands as a prominent contributor to cancer-related fatalities in women. Based on estimates from the World Health Organization (WHO) and the International Agency for Research on Cancer (IARC), there were approximately 529,000 new cases of cervical cancer worldwide in the year 2008. **Methods:** This study was done by retrieving information of 101 patients from records of Pathology Department in whom cervical malignancy was suspected by per speculum and per vaginal examination and confirmed by biopsy/hysterectomy. Standard streptavidin-biotin peroxidase method of IHC (immunohistochemical) was followed. **Results:** Increased expression of EGFR was seen in moderately differentiated squamous cell carcinoma than in adenocarcinoma and poorly differentiated SC carcinoma. P-value of EGFR over expression in MdSCC is <0.05, hence there is a significant association of EGFR over expression in MdSCC. **Conclusion:** The study revealed a noteworthy presence of the p16 biomarker in cervical cancer patients who were younger in age and at early stages of the disease. Consequently, the utilization of the p16 biomarker in screening or early diagnosis may contribute to improved prognosis for squamous cell carcinoma of the cervix.

**Keywords:** Cervical Cancer, EGFR, P16, Radiotherapy, Chemotherapy

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### INTRODUCTION

Cervical cancer is a prevalent and serious malignancy that poses a significant threat to women's health worldwide. In 2018, total 570,000 new cases are reported and 311,000 deaths. It stands as the fourth most commonly diagnosed cancer and the fourth leading cause of cancer-related deaths among women[1]. Notably, approximately 85% of cervical cancer deaths worldwide are concentrated in underdeveloped or developing countries, where the mortality rate is 18 times higher compared to wealthier nations[2]. In India, cervical cancer ranks as the second most common malignant tumor affecting women. [3-5].

In May 2018, the World Health Organization (WHO) made a global plea for the eradication of cervical

cancer. This call to action received a positive response from over 70 countries and international academic societies, who promptly initiated efforts in that direction[6, 7]. Subsequently, on November 17, 2020, WHO unveiled a comprehensive global strategy aimed at expediting the elimination of cervical cancer as a public health issue. This milestone marked the first collective commitment of 194 countries to eliminate cervical cancer[8, 9]. Multiple studies have consistently identified various risk factors associated with the development of cervical cancer. These factors include low economic status, inadequate personal and sexual hygiene, smoking, early onset of sexual activity, and engaging in multiple sexual partnerships[10, 11].

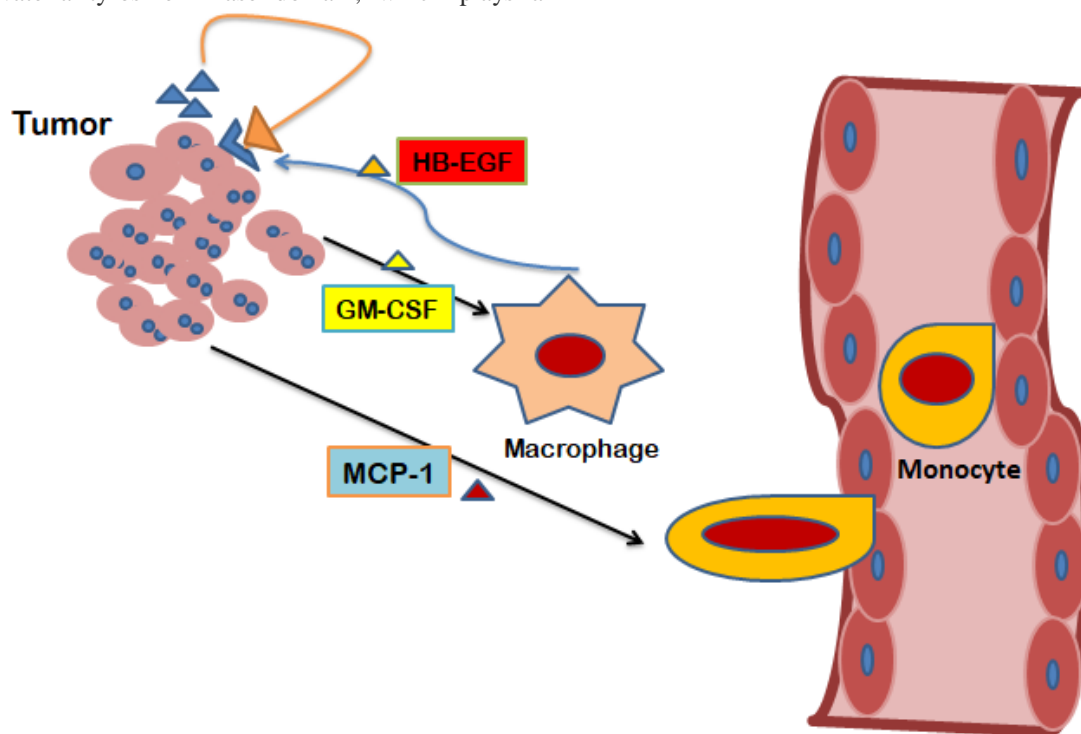
The primary causal factor in the process of carcinogenesis is human papillomavirus (HPV). However, it is important to note that not all HPV infections in women lead to cervical cancer. The progression from a normal cell to a precancerous lesion and eventually an invasive lesion is triggered by high-risk HPV genotypes. The pathogenesis of HPV infection involves the over expression of viral onco-proteins, which can hinder various cellular proteins and impact biological processes such as cell proliferation, the cell cycle, and apoptosis. The viral and host cellular changes that contribute to cervical carcinogenesis provide valuable insights into the nature of the disease and serve as a source of inspiration for the development of targeted molecular therapies[12-15].

Cervical cancer arises in the cervix, the narrow passage that connects the uterus to the vagina through the endocervical canal (Fig. 2) [16]. Early-stage cervical cancer is often asymptomatic and may be diagnosed during a routine screening or pelvic examination. The most common symptoms include heavy or abnormal vaginal bleeding, in particular following intercourse[17, 18].

Epidermal growth factor receptor (EGFR) is a trans-membrane glycoprotein receptor with a molecular weight of 170 kDa. It undergoes dimerization to activate a tyrosine kinase domain, which plays a

crucial role in regulating various cellular functions such as cell differentiation, growth, gene expression, and development[19, 20]. In cervical cancer, high expression of the epidermal growth factor receptor (EGFR) is associated with tumor development, as EGFR expression increases with increasing malignant stage and subsequent EGFR activation leads to cell growth, differentiation, resistance to apoptosis, cellcycle progression and angiogenesis. EGFR-based therapy has established efficacy in selected patients with head and neck squamous cell carcinoma, colorectal carcinoma and non-smallcell lung carcinoma[21-23].

The over expression of EGFR has been linked to decreased survival rates in patients with cervical cancer, and this relationship has been the subject of investigation for several years[24]. The effectiveness of EGFR inhibition as a treatment for cervical cancer is still uncertain. Although several clinical trials investigating EGFR inhibitors in cervical cancer patients have been conducted, the outcomes and impact of these drugs are not firmly established. It has been observed through numerous clinical trials that only a specific subgroup of patients shows a response to EGFR inhibitors. However, conflicting findings have been reported by various research groups. [25-27].



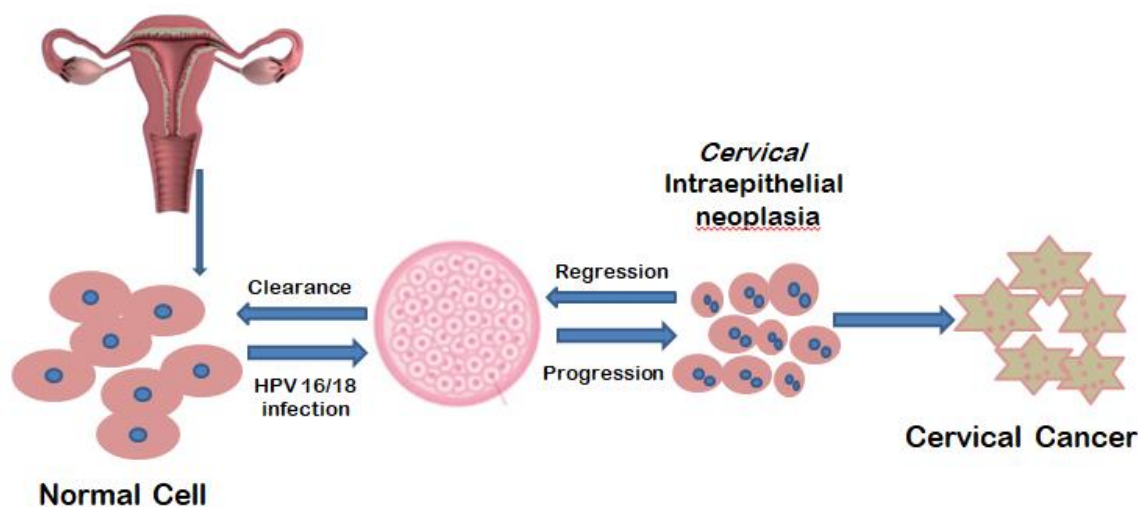
**Figure 1: Proposed mechanism of heparin-binding EGF-like growth factor (HB-EGF) expression and macrophage recruitment in cervical cancer patients**

It has been suggested that HB-EGF is expressed by cervical cancer-associated stromal fibroblasts, thus promoting cancer cell proliferation in a paracrine manner. In vitro experiments with head and neck squamous cell carcinoma cells show that this HB-EGF autocrine loop is associated with invasive processes

through the EGFR-Src-cortactin cascade[22, 28-30]. Cervical cancer cells are shown to express GM-CSF to attract macrophages to the tumor environment. Although macrophages and other stromal cells express the primary EGFR ligand HB-EGF, they do not appear to be the major source of HB-EGF, as the

relative amount of HB-EGF measured in the tumor cell compartment was on average four times higher

than the relative amount of HB-EGF in the stromal compartment (**Figure 1**)[22, 30, 31].



**Figure 2: Sequence of events leading to the development of HPV-related cervical carcinoma.**

Cervical intraepithelial neoplasia (CIN) occurs when squamous cells exhibit abnormal growth due to persistent HPV infection. While many of these changes naturally resolve over time, in some cases, the integration of viral particles leads to the transformation of cells and their progression into cervical cancer[30, 32]. As demonstrated in **Figure 2**, pre-cancerous changes can occur in cervix cells infected with high-risk HPV, and most of these changes resolve over time, but some do progress to cervical cancer. HPV is responsible for almost 100% of cervical cancers[30, 33].The genome of HPV encodes gynaecological cancer-related oncogenes, which include E5, E6 and E7. These oncogenes encode proteins that are implicated in carcinogenesis, a process that has been demonstrated in both in vitro and in vivo models[34, 35].

Most of HPV infections are asymptomatic and partners can unknowingly pass it on to one another through sexual contact. Majority of HPV-related cancers are prevented by using commercial HPV vaccines; however, these vaccines are ineffective in eradicating persistent HPV infections, and have not been demonstrated to slow down the HPV-related progression of malignant tumours [36-38]. To facilitate the development of cervical cancer, human papillomavirus (HPV) disrupts the cellular regulatory mechanisms by inducing the expression of viral proteins. These proteins, in turn, promote cellular transformation by modulating the expression of p53. It should be noted that the expression of p53 varies depending on the stage of cervical cancer[39, 40].

Cervical cancer cases exhibiting P16 expression have shown a more favorable prognosis, as reported in previous studies. High P16 expression in cervical cancer has been associated with a significant improvement in both five-year overall survival and disease-free survival rates. It is worth noting that the

present study has a limitation in terms of not conducting follow-up assessments to evaluate the long-term prognosis of the cases. Nevertheless, it was observed that P16 block positivity was particularly high among young females and this finding was statistically significant [41, 42]. The utilization of P16 as an additional biomarker has significantly enhanced diagnostic consensus. The expression of P16 is indicative of high-risk human papilloma virus (HR-HPV) infection and the integration of the viral genome into the host genome. However, it is important to note that different pathologists may employ varying criteria and thresholds for interpreting P16 expression[43, 44]. The Lower Anogenital Squamous Terminology (LAST) criteria categorize P16 immuno-reactivity into three groups: block positive, ambiguous, or negative. This classification is determined by evaluating the presence of P16 expression in the nucleus with or without cytoplasmic staining[45].

Further research is necessary for prognostic significance of EGFR and P16 in cervical cancer. In present study utilize immune-histochemical analysis of P16 and epidermal growth factor receptor (EGFR) to establish a diagnosis of cervical carcinoma in all cervical biopsies and hysterectomy specimens received.

## MATERIAL AND METHODS

A retrospective study was carried in the department of pathology at G.S.V.M. Medical College, Kanpur from January 2020 to January 2021 and all relevant clinical details and finding were revised on the requisition form. This study was done by retrieving information of 101 patients from records of Pathology Department in whom cervical malignancy was suspected by per speculum and per vaginal examination and confirmed by biopsy or hysterectomy. Standard streptavidin-

biotin peroxidase method of IHC (immunohistochemical) was followed [46, 47]. Sections were immunohistochemically stained for mouse monoclonal antibody to p16 manufactured by BioGenex p16 and EGFR. So, all adequate cervical biopsies and hysterectomy specimen of all age group were included in the study and Exclusion criteria for the study was patients who were already treated for cervical cancer. Inadequate biopsies.

**RESULT**

Total 101 case were included in this study, maximum case ( 57.14%) lie between the age group of 41-60 years. The mean age was 55 years. Well Differentiated carcinoma was the commonest grade. 40% of patients in this study was smokers suggesting that carcinoma of cervix were more common in smokers. Patients who have a history of use of contraceptive pills for contraception, shows higher grades of malignancy. However, duration of use was not taken into consideration.

It was observed that highest parity 5 was found in female with poorly differentiated SCC. Parity didn't have any significance effect on other grade of malignances.

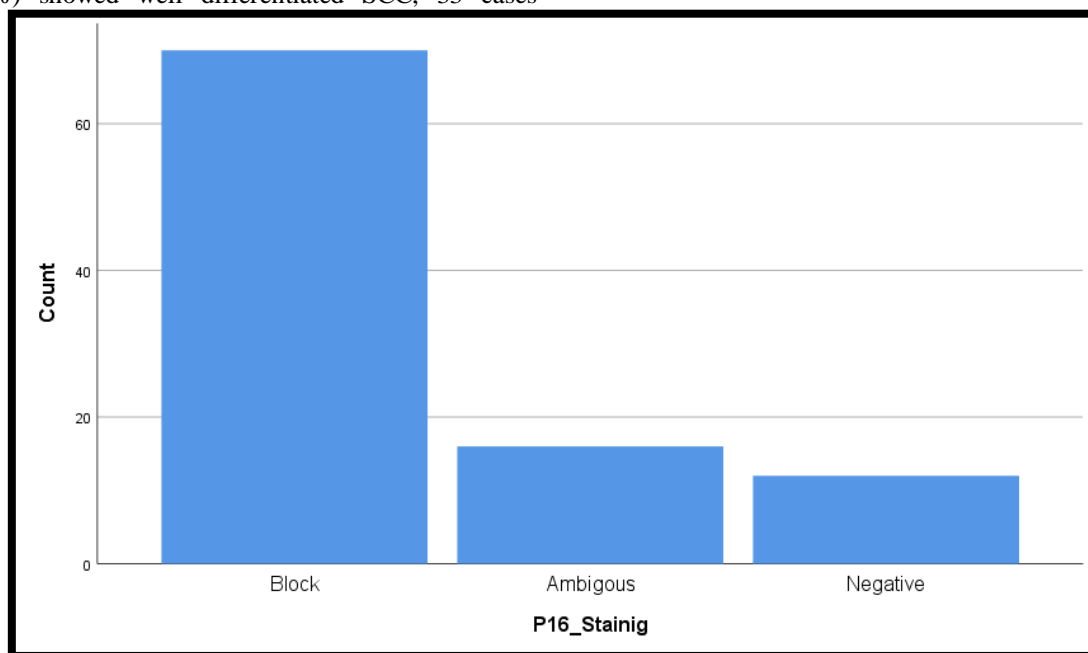
It was observed that carcinoma cervix was more common in postmenopausal women. A total of 114 cases (54%) were post-menopausal of which 24 cases (40%) showed well differentiated SCC, 33 cases

(57.90%) showed moderately differentiated SCC and 21 cases (58.33%) showed poorly differentiated SCC. It was also observed that carcinoma cervix was more common in women who gave history of irregular periods. The grade of malignancy did not show any specific relationship to menstrual status.

The well-differentiated squamous cell carcinoma exhibited diffuse block positivity when stained with P16 and well-differentiated squamous cell carcinoma, there was observed an over expression of EGFR in **figure 5 & 6** respectively.

Bleeding was comments symptom in all the cases with maximum frequency in poorly differentiated SCC (93%) followed by well differentiated SCC (75%). Lower abdominal pain was seen in a significant percentage of women with both well differentiated SCC and moderately differentiated SCC (85%), but slightly higher frequency of pain was found in women having adenocarcinoma (87%). Discharge per vaginal was seen only in a small percentage cases, mostly in poorly differentiated malignancy. The other findings on examination in order of frequency were bleeding on touch and necrosis. The presentation did not alter with the grade of malignancy.

In the current study the total number of cases showing P16 block, ambiguous and no positivity were 70, 13 and 9 respectively as shown in **Figure 3** below.



**Figure 3: Showing P16 staining pattern in cervical carcinoma cases**

It was observed that increased block positivity of p16 was seen in moderately differentiated squamous cell carcinoma than in adenocarcinoma and poorly differentiated carcinoma SCC. P-value of p16 block staining in MdSCC is <0.05, hence there is a significant association of p16 block positivity in MdSCC as shown in **Table 1 & 2**.

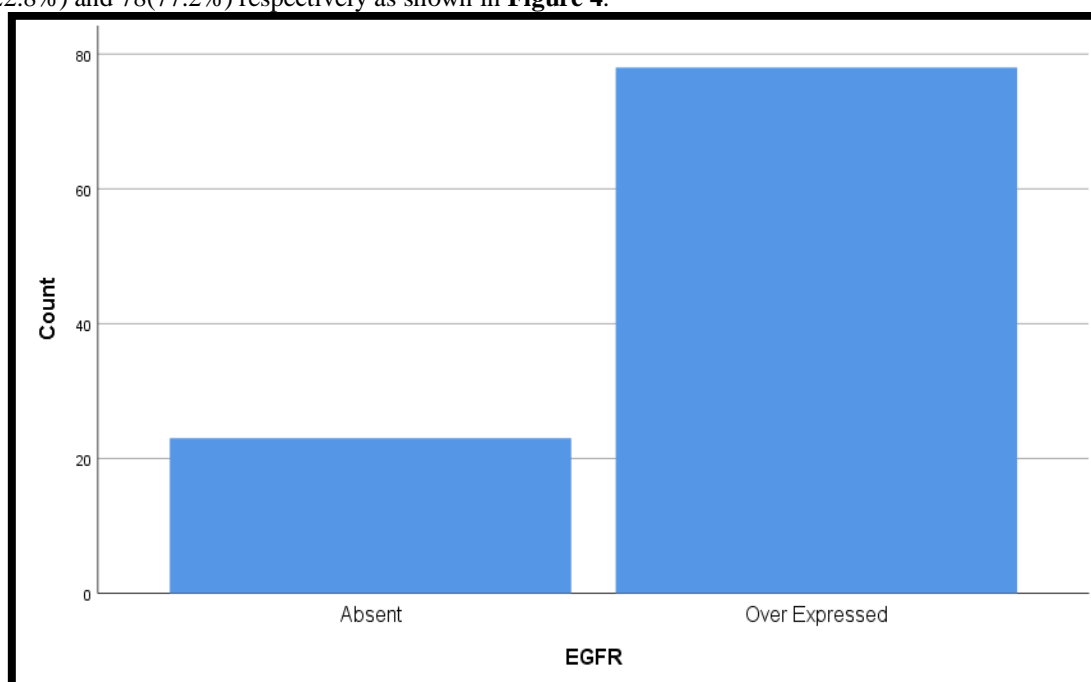
**Table1: Showing correlation of intensity of P16stainingwithWdSCC & MdSCC**

			WdSCC			MdSCC		
			No	Yes	Total	No	Yes	Total
P16_Stainig	Block	Count	52	18	70	23	47	70
		% within P16_Stainig	74.3	25.7	100	32.9	67.1	100
	Ambiguous	Count	14	2	16	8	8	16
		% within P16_Stainig	87.5	12.5	100	50.0	50.0	100
	Negative	Count	8	4	12	9	3	12
		% within P16_Stainig	66.7	33.3	100	75.0	25.0	100
Total		Count	74	24	98	40	58	98
		% within P16_Stainig	75.5	24.5	100	40.8	59.2	100

**Table2: Showing correlation of intensity of P16 Staining with PdSCC & Adeno CA**

			Poorly differentiated SCC			Adeno CA		
			No	Yes	Total	No	Yes	Total
P16_Stainig	Block	Count	65	5	70	70	0	70
		% within P16_Stainig	92.9	7.1	100	100	0.0	100
	Ambiguous	Count	13	3	16	13	3	16
		% within P16_Stainig	81.3	18.8	100	81.3	18.8	100
	Negative	Count	9	3	12	10	2	12
		% within P16_Stainig	75.0	25.0	100	83.3	16.7	100
Total		Count	87	11	98	93	5	98
		% within P16_Stainig	88.8	11.2	100	4.0	5.1	100

In the current study the total number of cases showing EGFR expression as absent and over expression are 23(22.8%) and 78(77.2%) respectively as shown in **Figure 4**.



**Figure 4: Showing EGFR expression in total number of cases**

It was observed that increased expression of EGFR was seen in moderately differentiated squamous cell carcinoma than in adenocarcinoma and poorly differentiated SC carcinoma. P-value of EGFR over expression in MdSCC is <0.05, hence there is a significant association of EGFR over expression in MdSCC as shown in **Table3 &4**.



**Table3: Showing correlation of EGFR expression with WdSCC & MdSCC**

			WdSCC		Total	MdSCC		Total
			No	Yes		No	Yes	
EGFR	Absent	Count	16	7	23	17	6	23
		% within EGFR	69.6	30.4	100	73.9	26.1	100
	Over Expressed	Count	58	20	78	26	52	78
		% within EGFR	74.4	25.6	100	33.3	66.7	100
Total		Count	74	27	101	43	58	101
		% within EGFR	73.3	26.7	100	42.6	57.4	100

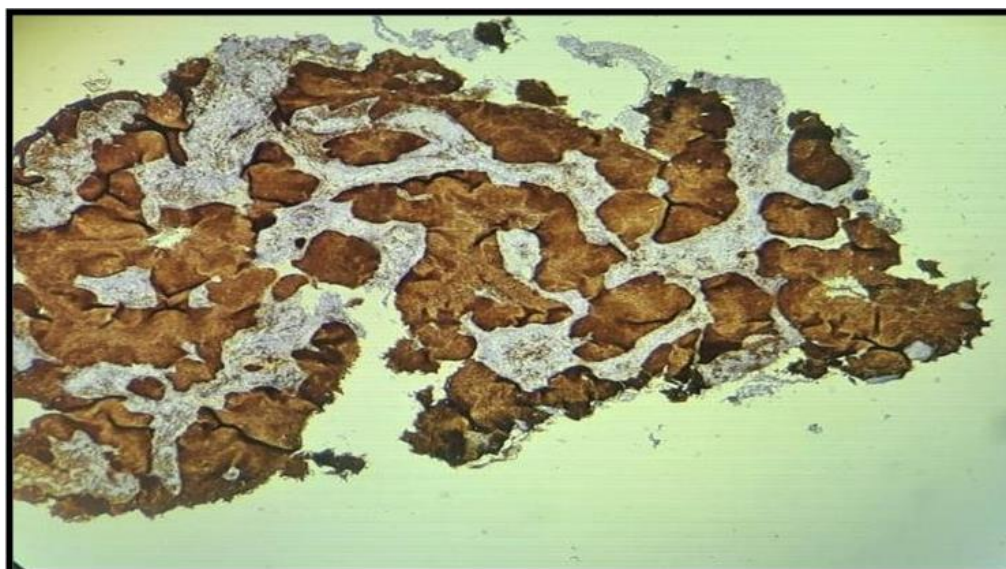
**Table4: Showing correlation of expression of EGFR with PDSCC**

			Poorly differentiated SCC		Total	Adenocarcinoma		Total
			No	Yes		No	Yes	
EGFR	Absent	Count	16	7	23	20	3	23
		% within EGFR	69.6	30.4	100	87	13	100
	Over Expressed	Count	74	4	78	76	02	78
		% within EGFR	94.9	5.1	100	97.4	2.6	100
Total		Count	90	11	101	96	5	101
		% within EGFR	89.1	10.9	100	95	5	100

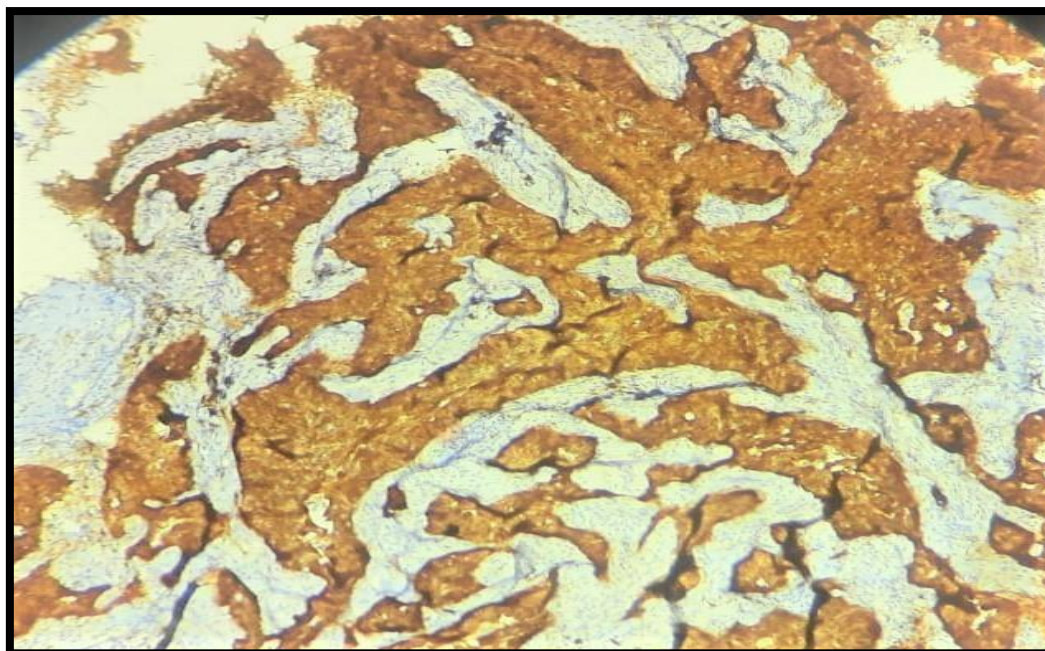
In the current study there is no significant correlation between P16 and EGFR especially in squamous cell carcinoma cases as shown in **Table 5**.

**Table5: Showing correlation of P16stainingwithEGFR**

			EGFR		Total
			Absent	Over Expressed	
P16_Stainig	Block	Count	10	60	70
		% withinP16_Stainig	14.3%	85.7%	100.0%
	Ambiguous	Count	6	10	16
		% withinP16_Stainig	37.5%	62.5%	100.0%
	Negative	Count	6	6	12
		% withinP16_Stainig	50.0%	50.0%	100.0%
Total		Count	22	76	98
		% withinP16_Stainig	22.4%	77.6%	100.0%



**Figure5: Showing P16 staining in well differentiated squamous cell Carcinoma showing diffuse block positivity(IHC; 400X magnification)**



**Figure 6: EGFR over expression in well differentiated Squamous cell carcinoma (IHC; 400 X magnifications).**

#### DISCUSSION

Cervical malignancy is the 2<sup>nd</sup> most common malignancy of the female[48]. This retrospective study was done in the department of pathology; GSVM Medical College, Kanpur and a total of 101 cases were involved in this study. In this study it was observed that mean age for cervical carcinoma was  $\geq 55$  years and region for higher incidence of cervical carcinoma in this age group were effect of hormonal change and increase susceptibility to genital tract infection. Strategies to prevent cervical cancer, both primary and secondary, play a crucial role in reducing the impact of this disease. There has been extensive literature addressing these prevention strategies [49, 50].

In some cases, women may exhibit a vaginal discharge that can vary in consistency from watery to mucoid or purulent, accompanied by an unpleasant odor. However, it is important to note that this symptom is rarely observed in isolation and is often accompanied by other symptoms. In advanced stages of the disease, patients may experience additional symptoms such as lower limb edema, flank pain, as well as pelvic or lower back pain[51]. In patients exhibiting any symptoms of cervical cancer, a pelvic examination is conducted, which includes the visualization of the cervix and vaginal mucosa. If any abnormalities are observed, a biopsy is performed[52].

Radiotherapy, which involves the use of high-energy X-rays, is a primary treatment modality in the management of cervical cancer[53]. Chemotherapy plays a crucial role in the standard treatment protocol for cervical cancer. It is commonly used as an adjuvant therapy following surgery, particularly when poor prognostic tumor characteristics elevate the risk

of disease recurrence. Additionally, chemotherapy is administered in combination with radiotherapy, as mentioned earlier, and as a standalone treatment for cases of locally advanced disease[54].

The use of immunotherapy, specifically targeting HPV oncoproteins, has emerged as a promising approach in the treatment of cervical cancer. This novel treatment method offers the advantage of specifically targeting dysplastic precancerous and malignant cervical epithelial cells that express HPV oncoproteins [55].

Chemotherapy agents are known to have a cytotoxic effect on both cancer cells and normal rapidly dividing cells, leading to various side effects such as anemia and alopecia. In contrast, targeted therapies are designed to specifically inhibit molecules, typically proteins, that are specifically expressed by cancer cells. These targeted therapies aim to disrupt the growth, proliferation, and metastasis of cancer cells[56].

As targeted therapies have a higher specificity for cancer cells compared to normal cells, it is expected that they will exhibit increased efficacy and reduced adverse effects compared to current chemotherapies. This anticipation stems from the fact that targeted therapies selectively target cancer cells, resulting in a more precise and focused treatment approach[57].

In the current study it was observed that highest parity 5 was found in female with Poorly Differentiated Squamous cell carcinoma and that parity have significance role in cervical carcinoma which is in accordance to the study of Natthakan Vanakankovitetal(2008)[58]. In our study it was observed that carcinoma cervix was more common in most postmenopausal women. It was also observed that carcinoma cervix more common in women who

gave history of irregular periods which was quiet similar to the study of Gloria YFetal(1998)[59].

In our study also shows bleeding was presented in all the cases with maximum frequency in poorly differentiated Squamous cell carcinoma (93%) followed by well differentiated Squamous cell carcinoma (75%) lower abdominal pain in significant percentage of women which is similar to the study of Louise A et al (1987)[60]. It was observed that mean count increased with increasing grade of malignancy. Lowest count obtained in *In Situ* Squamous cell carcinoma to Moderately Differentiated Squamous cell carcinoma and higher in poorly differentiated Squamous cell carcinoma which is similar to the results observed by Singh Uma et al (2006)[61].

In the current study, increased block positivity of p16 was seen in moderately differentiated squamous cell carcinoma in comparison to adenocarcinoma and poorly differentiated carcinoma. P-value of p16 staining in MdSCC is <0.05, hence there is a significant association of p16 block positivity in MdSCC.

Kalyani et al (2020) showed that there is significant association between age and p16 expression, where all cases between 30-59 years showed block positivity. Among the post-menopausal women, women between 40-44 yrs showed maximum block positivity (93.75%)[62]. Sarwath et al (2017) stated that there was a significant correlation between p16 expression and age group between 41-60 years. Hence, p16 is an appropriate surrogate marker for use in early screening of cervical cancer [63]. Fu HC et al (2018) stated that p16 expression was not found to have an association with tumor stage, tumor size, histological grade, vascular invasion[42].

T. Soonthorn thumetal(2011) stated that EGFR over expression is associated with poor prognostic factors such as increased tumor size, lymphnode metastases and recurrence of disease. The average percentage of squamous cell carcinoma over expressing EGFR is 51% compared with 23% for adenocarcinoma and 38% for adenosquamous carcinoma[64]. According to D Pfeiffer et al (1998), who suggested that EGFR expression may be indicative of the biological aggressiveness of cervical squamous cell carcinoma[65].

In the current study, we found that compared to EGRF expression in squamous cell carcinoma versus adenocarcinoma and poorly differentiated carcinoma, the squamous cell type always stains more strongly, that means strong positivity for squamous cell carcinoma as compared to other type of carcinoma like adenocarcinoma.

## CONCLUSION

The study revealed a noteworthy presence of the p16 biomarker in cervical cancer patients who were younger in age and at early stages of the disease. Consequently, the utilization of the p16 biomarker in screening or early diagnosis may contribute to improved prognosis for squamous cell carcinoma of

the cervix. Additionally, this finding suggests the potential application of targeted therapies based on the anti-cancer properties of the p16 protein. The importance of p16 in cervix is that it is seen maximum in moderate grade lesion of SCC more than any other type of SCC or adenocarcinoma. Therefore, p16 biomarker may have a beneficial use in screening or early diagnosis leading to better prognosis of squamous cell carcinoma of cervix. It has been showed that squamous cell carcinoma of the cervix frequently express EGFR, often at high concentration and least frequently in adenocarcinoma. Current study showed that immuno-histochemical staining of elevated expression of EGF receptor in cervical squamous epithelium was associated with its neoplastic transformation and emerged as a tumor marker for cervical squamous neoplasia.

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## CONFLICT OF INTEREST

All the authors declare that they have no conflict of interest.

## ETHICAL STATEMENT

All the necessary permissions are obtained from the institution.

## AUTHOR'S CONTRIBUTIONS

Anjana Trivedi: Concept, Study design, data collection, literature search, statistical analysis, manuscript writing. Vandana Mishra Tiwari: Manuscript writing and editing, Dr. Suman Lata Verma: manuscript editing, Anita Omhare: manuscript review

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Nil

## ABBREVIATIONS

DFS: Disease Free Survival  
 HPV: Human Papilloma Virus  
 IHC: Immunohistochemistry  
 MdSCC: Moderately Differentiated Squamous Cell Carcinoma  
 SCC: Squamous Cell Carcinoma  
 WdSCC: Well Differentiated Squamous Cell Carcinoma  
 EGFR: Epidermal growth factor receptor  
 HB-EGF: Heparin-binding epidermal growth factor  
 IARC: International Agency for Research on Cancer  
 CIN: Cervical intraepithelial Neoplasia

## REFERENCES

- Zhang, X., et al., Trends of cervical cancer at global, regional, and national level: data from the Global Burden of Disease study 2019. BMC public health, 2021. 21(1): p. 1-10.



2. Torre, L.A., et al., Global cancer statistics, 2012. *CA Cancer J Clin*, 2015. 65(2): p. 87-108.
3. Chen, J., et al., Toripalimab combined with concurrent platinum-based Chemoradiotherapy in patients with locally advanced cervical Cancer: an open-label, single-arm, phase II trial. *BMC cancer*, 2022. 22(1): p. 1-9.
4. Hategeka, C., et al., Implementation research on noncommunicable disease prevention and control interventions in low-and middle-income countries: A systematic review. *PLoS Medicine*, 2022. 19(7): p. e1004055.
5. Amini, M., F. Zayeri, and M. Salehi, Trend analysis of cardiovascular disease mortality, incidence, and mortality-to-incidence ratio: results from global burden of disease study 2017. *BMC Public Health*, 2021. 21(1): p. 1-12.
6. Organization, W.H., General meeting of the WHO global coordination mechanism on the prevention and control of noncommunicable diseases: meeting report: International Conference Centre, Geneva, Switzerland, 5-6 November 2018. 2019, World Health Organization.
7. Alyanak, O., Turkey's Diyanet and Political Islam during the Pandemic. *Viral Loads: Anthropologies of Urgency in the Time of COVID-19*, ed. Lenore Manderson, Nancy J. Burke, Ayo Wahlberg,(London: UCL Press, 2021), 2021: p. 162-180.
8. Organization, W.H., Programme Budget 2018–2019: Implementation and mid-term review. 2019, World Health Organization. Regional Office for South-East Asia.
9. Garland, S.M., et al., IPVS policy statement on HPV nucleic acid testing guidance for those utilising/considering HPV as primary precancer screening: Quality assurance and quality control issues. *Journal of Clinical Virology*, 2023. 159: p. 105349.
10. Chan, C.K., et al., Human papillomavirus infection and cervical cancer: epidemiology, screening, and vaccination—review of current perspectives. *Journal of oncology*, 2019. 2019.
11. Kashyap, N., et al., Risk factors of cervical cancer: a case-control study. *Asia-Pacific journal of oncology nursing*, 2019. 6(3): p. 308-314.
12. Tjalma, W., et al., Role of human papillomavirus in the carcinogenesis of squamous cell carcinoma and adenocarcinoma of the cervix. *Best practice & research Clinical obstetrics & gynaecology*, 2005. 19(4): p. 469-483.
13. Burd, E.M., Human papillomavirus and cervical cancer. *Clinical microbiology reviews*, 2003. 16(1): p. 1-17.
14. Balasubramaniam, S.D., et al., Key molecular events in cervical cancer development. *Medicina*, 2019. 55(7): p. 384.
15. Boyle, P., et al., Epidemiology of mouth cancer in 1989: a review. *J R Soc Med*, 1990. 83(11): p. 724-30.
16. SHUKLA, D., CERVICAL CANCER: FACTS EVERY WOMEN NEEDS TO KNOW.
17. Thistlethwaite, J. and R. Stewart, Routine pelvic examination for asymptomatic women: exploring the evidence. *Australian family physician*, 2006. 35(11).
18. Mwaka, A.D., et al., Symptomatic presentation with cervical cancer in Uganda: a qualitative study assessing the pathways to diagnosis in a low-income country. *BMC women's health*, 2015. 15(1): p. 1-13.
19. Huerta, J., et al., Epidermal growth factor receptor in adult human dorsal root ganglia. *Anatomy and embryology*, 1996. 194: p. 253-257.
20. Schaeffer, H.J. and M.J. Weber, Mitogen-activated protein kinases: specific messages from ubiquitous messengers. *Molecular and cellular biology*, 1999. 19(4): p. 2435-2444.
21. Cheng, W.-L., et al., The role of EREG/EGFR pathway in tumor progression. *International journal of molecular sciences*, 2021. 22(23): p. 12828.
22. Schrevel, M., et al., Autocrine expression of the epidermal growth factor receptor ligand heparin-binding EGF-like growth factor in cervical cancer. *International Journal of Oncology*, 2017. 50(6): p. 1947-1954.
23. Dewi, I.G.A.S.M., N.P. Sriwidayanti, and N.P. Ekawati, The role of epidermal growth factor receptor as progression factor in cervical intraepithelial neoplasia and squamous cell carcinoma. *Bali Medical Journal*, 2021. 10(1): p. 238-242.
24. Tian, W.-J., et al., Prognostic impact of epidermal growth factor receptor overexpression in patients with cervical cancer: a meta-analysis. *PLoS One*, 2016. 11(7): p. e0158787.
25. Narayanan, R., et al., Epidermal growth factor-stimulated human cervical cancer cell growth is associated with EGFR and cyclin D1 activation, independent of COX-2 expression levels. *International journal of oncology*, 2012. 40(1): p. 13-20.
26. Van Kessel, K.E., et al., Targeted therapies in bladder cancer: an overview of in vivo research. *Nature reviews Urology*, 2015. 12(12): p. 681-694.
27. Eberhard, D.A., G. Giaccone, and B.E. Johnson, Biomarkers of response to epidermal growth factor receptor inhibitors in Non-Small-Cell Lung Cancer Working Group: Standardization for use in the clinical trial setting. *Journal of Clinical Oncology*, 2008. 26(6): p. 983-994.
28. Murata, T., et al., HB-EGF and PDGF mediate reciprocal interactions of carcinoma cells with cancer-associated fibroblasts to support progression of uterine cervical cancers. *Cancer research*, 2011. 71(21): p. 6633-6642.
29. Schrevel, M., et al., Molecular mechanisms of epidermal growth factor receptor overexpression in patients with cervical cancer. *Modern Pathology*, 2011. 24(5): p. 720-728.
30. Makgoo, L., S. Mosebi, and Z. Mbita, Molecular Mechanisms of HIV Protease Inhibitors Against HPV-Associated Cervical Cancer: Restoration of TP53 Tumour Suppressor Activities. *Frontiers in Molecular Biosciences*, 2022. 9.
31. Rigo, A., et al., Macrophages may promote cancer growth via a GM-CSF/HB-EGF paracrine loop that is enhanced by CXCL12. *Molecular cancer*, 2010. 9: p. 1-13.
32. Baak, J.P., et al., Dynamic behavioural interpretation of cervical intraepithelial neoplasia with molecular biomarkers. *Journal of clinical pathology*, 2006. 59(10): p. 1017-1028.
33. Szymonowicz, K.A. and J. Chen, Biological and clinical aspects of HPV-related cancers. *Cancer biology & medicine*, 2020. 17(4): p. 864.
34. Hemmat, N. and H. Bannazadeh Baghi, Association of human papillomavirus infection and inflammation in cervical cancer. *Pathogens and disease*, 2019. 77(5): p. ftz048.

35. Schmitz, M., et al., Non-random integration of the HPV genome in cervical cancer. *PloS one*, 2012. 7(6): p. e39632.
36. McPartland, T.S., et al., Men's perceptions and knowledge of human papillomavirus (HPV) infection and cervical cancer. *Journal of American College Health*, 2005. 53(5): p. 225-230.
37. Hansen, B.T., S. Campbell, and M. Nygård, Long-term incidence trends of HPV-related cancers, and cases preventable by HPV vaccination: a registry-based study in Norway. *Bmj Open*, 2018. 8(2): p. e019005.
38. Vici, P., et al., Immunologic treatments for precancerous lesions and uterine cervical cancer. *Journal of Experimental & Clinical Cancer Research*, 2014. 33: p. 1-15.
39. Song, D., et al., Effect of human papillomavirus infection on the immune system and its role in the course of cervical cancer. *Oncology letters*, 2015. 10(2): p. 600-606.
40. Sato, Y. and T. Tsurumi, Genome guardian p53 and viral infections. *Reviews in medical virology*, 2013. 23(4): p. 213-220.
41. Nicolás, I., et al., Prognostic implications of genotyping and p16 immunostaining in HPV-positive tumors of the uterine cervix. *Modern Pathology*, 2020. 33(1): p. 128-137.
42. Fu, H.-C., et al., Low P16INK4A expression associated with high expression of cancer stem cell markers predicts poor prognosis in cervical cancer after radiotherapy. *International Journal of Molecular Sciences*, 2018. 19(9): p. 2541.
43. Cuschieri, K. and N. Wentzensen, Human papillomavirus mRNA and p16 detection as biomarkers for the improved diagnosis of cervical neoplasia. *Cancer Epidemiology Biomarkers & Prevention*, 2008. 17(10): p. 2536-2545.
44. Nemes, J.A., et al., Expression of p16INK4A, p53, and Rb proteins are independent from the presence of human papillomavirus genes in oral squamous cell carcinoma. *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontology*, 2006. 102(3): p. 344-352.
45. Liu, Y., et al., Using p16 immunohistochemistry to classify morphologic cervical intraepithelial neoplasia 2: correlation of ambiguous staining patterns with HPV subtypes and clinical outcome. *Human pathology*, 2017. 66: p. 144-151.
46. Pileri, S.A., et al., Antigen retrieval techniques in immunohistochemistry: comparison of different methods. *The Journal of Pathology: A Journal of the Pathological Society of Great Britain and Ireland*, 1997. 183(1): p. 116-123.
47. Bussolati, G., et al., Retrieved endogenous biotin: a novel marker and a potential pitfall in diagnostic immunohistochemistry. *Histopathology*, 1997. 31(5): p. 400-407.
48. Abdulkareem, F. Epidemiology and incidence of common cancers in Nigeria. in Paper presented. 2009.
49. Gupta, A.K., et al., Oral Cancer Risk Assessment for Different Types of Smokeless Tobacco Products Sold Worldwide: A Review of Reviews and Meta-analyses. *Cancer Prev Res (Phila)*, 2022. 15(11): p. 733-746.
50. Rock, C.L., et al., Prevention of cervix cancer. *Critical reviews in oncology/hematology*, 2000. 33(3): p. 169-185.
51. Mylonas, I. and F. Bergauer, Diagnosis of vaginal discharge by wet mount microscopy: a simple and underrated method. *Obstetrical & Gynecological Survey*, 2011. 66(6): p. 359-368.
52. Jordan, J., et al., European guidelines for quality assurance in cervical cancer screening: recommendations for clinical management of abnormal cervical cytology, part 1. *Cytopathology*, 2008. 19(6): p. 342-354.
53. Skliarenko, J. and P. Warde, Radiotherapy: practical applications and clinical aspects. *Medicine*, 2011. 39(12): p. 705-710.
54. Movva, S., et al., Novel chemotherapy approaches for cervical cancer. *Cancer: Interdisciplinary International Journal of the American Cancer Society*, 2009. 115(14): p. 3166-3180.
55. Peralta-Zaragoza, O., et al., Targeted treatments for cervical cancer: a review. *OncoTargets and therapy*, 2012: p. 315-328.
56. Thirumaran, R., G.C. Prendergast, and P.B. Gilman, Cytotoxic chemotherapy in clinical treatment of cancer, in *Cancer immunotherapy*. 2007, Elsevier. p. 101-116.
57. Senapati, S., et al., Controlled drug delivery vehicles for cancer treatment and their performance. *Signal transduction and targeted therapy*, 2018. 3(1): p. 7.
58. Vanakankovit, N. and S. Taneepanichskul, Effect of oral contraceptives on risk of cervical cancer. *Medical journal of the Medical Association of Thailand*, 2008. 91(1): p. 7.
59. Ho, G.Y., et al., Natural history of cervicovaginal papillomavirus infection in young women. *New England Journal of Medicine*, 1998. 338(7): p. 423-428.
60. Brinton, L.A., et al., Sexual and reproductive risk factors for invasive squamous cell cervical cancer. *Journal of the National Cancer Institute*, 1987. 79(1): p. 23-30.
61. Misra, J.S. and U. Singh, Results of longterm hospital based cytological screening in asymptomatic women. *Diagnostic Cytopathology*, 2006. 34(3): p. 184-187.
62. Kalyani, R., C. Raghuv eer, and S. Sheela, Expression of P16 biomarker in squamous cell carcinoma of uterine cervix and its association with clinicopathological parameters: A cross-sectional study. *Biomedical Research and Therapy*, 2020. 7(9): p. 3954-3961.
63. Sarwath, H., et al., Introduction of p16INK4a as a surrogate biomarker for HPV in women with invasive cervical cancer in Sudan. *Infectious agents and cancer*, 2017. 12(1): p. 1-8.
64. Soonthornthum, T., et al., Epidermal growth factor receptor as a biomarker for cervical cancer. *Annals of oncology*, 2011. 22(10): p. 2166-2178.
65. Pfeiffer, D., et al., Epidermal-growth-factor receptor correlates negatively with cell density in cervical squamous epithelium and is down-regulated in cancers of the human uterus. *International journal of cancer*, 1998. 79(1): p. 49-55.